

Phenolic Hydrazones Are Potent Inhibitors of Macrophage Migration Inhibitory Factor Proinflammatory Activity and Survival Improving Agents in Sepsis

Darrin R. Dabideen,[†] Kai Fan Cheng,[†] Bayan Aljabari,[†] Edmund J. Miller,[‡] Valentin A. Pavlov,[§] and Yousef Al-Abed^{*,†,‡,#}

Laboratory of Medicinal Chemistry, Laboratory of Cardiopulmonary Research, and Laboratory of Biomedical Sciences, The Feinstein Institute for Medical Research, 350 Community Drive, Manhasset, New York 11030, and The New York School of Medicine, New York, New York 10016

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A series of phenolic hydrazones were synthesized and evaluated for their inhibition of macrophage migration inhibitory factor (MIF) tautomerase activity. Compound **7** emerged as a potent inhibitor of MIF with an IC₅₀ of 130 nM. Compound **7** dose-dependently suppressed TNF α secretion from lipopolysaccharide stimulated macrophages. The therapeutic importance of the MIF inhibition by **7** is demonstrated by the significant protection from the lethality of sepsis when administration of the compound was initiated in a clinically relevant time frame.

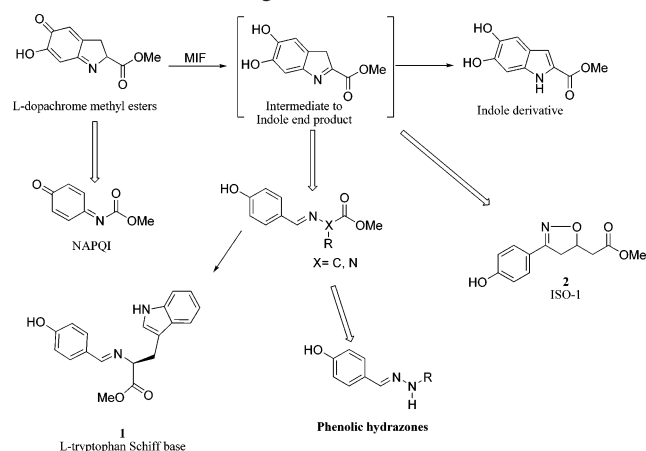
Introduction

Macrophage migration inhibitory factor (MIF^α) is a potent proinflammatory cytokine critically involved in the pathogenesis of sepsis and other inflammatory disorders.^{1,2} We and others have demonstrated that MIF is an important late-acting mediator of systemic inflammation and that inhibition of its activity in vivo attenuates the lethal consequences of endotoxemia and sepsis in rodents.^{3,4}

MIF exists as a homotrimer^{5–8} with the unique ability to catalyze the tautomerization of nonphysiological substrates such as D-dopachrome and L-dopachrome methyl ester into their respective indole derivatives (Scheme 1).⁹ While the physiological role of the tautomerase activity is uncertain, we have previously found that compounds that are structurally similar to D- and L-dopachrome could bind to and thereby block the MIF's tautomerase active site.^{4,10–13} We have previously shown that NAPQI forms a covalent complex with MIF at its active site (Scheme 1) and is capable of irreversibly inhibiting the adverse biological effect of MIF.¹³ However, the toxicity of NAPQI precluded its use as a viable clinical inhibitor of MIF and the development of nontoxic small-molecule inhibitors of MIF tautomerase activity warranted further investigation.

The indole intermediate of MIF tautomerase catalysis presented itself as a suitable template for the development of potential MIF inhibitors. We reasoned that compounds designed around the phenylimine scaffold could act as potential MIF antagonists. Indeed, from our rational design we developed the amino acid Schiff bases and the isoxazoline compounds as MIF tautomerase inhibitors.^{10,12} Among the amino acid Schiff bases tested for their ability to inhibit the tautomerase activity of MIF, it was found that **1** was the most potent (Scheme 1, Table 1). Recently, we have reported an isoxazoline inhibitor of MIF, **2**

Scheme 1. Rational Design of MIF Inhibitors



(ISO-1), which blocks the tautomerase site, inhibits the ability of MIF to overcome anti-inflammatory glucocorticoid activities in vitro, and improves survival in animal models of experimental sepsis (Scheme 1).^{4,12}

In our quest to find more potent inhibitors of MIF, we revisited the phenylimine scaffold and modified it by adding nitrogen to afford the hydrazone type compound (Scheme 1). Our previous studies on the amino acid Schiff bases and compound **2** revealed that a para hydroxyl group, as exemplified by a phenolic moiety, is a key structural feature and is required for activity. We have shown that replacing the phenolic moieties of **1** and **2** with phenyl or halide-substituted phenyl or *p*-methoxyphenyl groups resulted in the decrease or complete loss of their ability to inhibit MIF tautomerase activity.^{10,12} In accordance with this result, the chemical structures of the new pharmacophores must contain a phenolic ring (Scheme 1). Utilizing this key structural feature and the observations of **1** and **2** complexed with MIF, we propose the synthesis of phenolic hydrazones **3–15** as potential MIF inhibitors.

Results and Discussion

The phenolic hydrazones were obtained in high yields by condensing *p*-hydroxybenzaldehyde and the appropriate hydrazone in an alcoholic solvent in the presence of an acid catalyst at room temperature. The hydrazones obtained were assayed

* To whom correspondence should be addressed. Phone: 516 562 3406. Fax: 516 562 1022. E-mail: yalabed@nshs.edu.

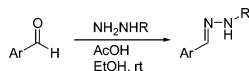
[†] Laboratory of Medicinal Chemistry, The Feinstein Institute for Medical Research.

[‡] Laboratory of Cardiopulmonary Research, The Feinstein Institute for Medical Research.

[§] Laboratory of Biomedical Sciences, The Feinstein Institute for Medical Research.

[#] The New York School of Medicine.

^α Abbreviations: MIF, macrophage migration inhibitory factor; NAPQI, *N*-acetyl-*p*-benzoquinone imine; LPS, lipopolysaccharide; CLP, cecal ligation and puncture.

Table 1. Phenolic Hydrazones and IC₅₀ for Inhibition of MIF Tautomerase Activity

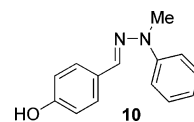
Compounds ^a	Ar	R	IC ₅₀ (μM) ^b
1 L-tryptophan Schiff base			1.6
2 ISO-1			7
3		H	>500
4		CH ₃	43
5			2.6
6^c			0.48
7^c			0.13
8^c			0.22
9^c			0.33

^a Compounds were characterized by ¹H NMR, ¹³C NMR, and MS.

^b Spectrophotometric analysis of MIF tautomerase activity on L-dopachrome methyl ester (see Experimental Section). ^c See Experimental Section for general procedure for the synthesis of **6–9**.

for their potency of inhibiting MIF tautomerase activity (Table 1).

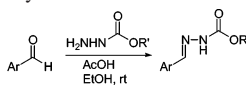
All of the hydrazones prepared in this study **3–15** have one common key structural feature; they all possess a “phenolic head” in the form of a 4-hydroxyphenyl ring. We and other workers have shown that the phenolic ring forms key hydrogen bond interactions between the amino acid residue asparagine-97C of the hydrophobic surface within the MIF active site.^{10,12,14} In addition to this important hydrogen bond interaction, there is a hydrophobic interaction that exists between the aromatic ring of the phenol and the side chains of the amino acid residues Pro-1, Met-2, Ile-64, Tyr-95, Val-106, and Phe-113 of the hydrophobic pocket that further contributes to the binding of the inhibitor.¹⁴ Supporting evidence for the key interaction between the hydroxyl functionality and the amino acid residue asparagine-97C was obtained by (1) modifying its position and (2) replacing the hydroxyl group with other functional groups. The position of the hydroxyl group is a critical feature of the hydrazones in that changing its position from para (IC₅₀ = 2.5 μM) to meta (IC₅₀ = 150 μM) resulted in a dramatic loss of its ability to inhibit MIF tautomerase activity. Furthermore, replacing the hydroxyl group with hydrogen, fluoro, amino, methoxy, and nitro functionalities afforded hydrazones that were inactive (data not shown). In developing a structure–activity relationship between the phenolic hydrazones and MIF, we synthesized hydrazones **3–6** from the simple hydrazine, methylhydrazine, phenylhydrazine, and *p*-methoxyphenylhydrazine. One further prerequisite for activity of our phenolic hydrazones is that they require a hydrophobic tail. This is evident from **3** (IC₅₀ ≥ 500 μM). The hydrogen substituent of the hydrazone in this compound does not offer any hydrophobic interactions and as








**Figure 1.** Disubstituted hydrazone.

a result is a very poor inhibitor of MIF tautomerase activity (Table 1). Upon replacement of the hydrogen with a methyl group, a pronounced improvement in the ability of the methylhydrazone **4** to inhibit the tautomerase activity of MIF (IC₅₀ = 43 μM) was observed. This suggests that there is a hydrophobic interaction between the methyl group and the surface of MIF. Replacing the methyl group of **4** with the more hydrophobic phenyl ring affords the hydrazone **5** (IC₅₀ = 2.6 μM) that is 16 times more potent. Installation of a *p*-methoxy group on the aromatic ring of **5** gives hydrazone **6** that is 5-fold more potent as an inhibitor than the parent compound. This increased binding of the *p*-methoxyphenylhydrazone may be explained by hydrogen bond interactions between the ether oxygen and the known amino acid residues and by the possibility of π–π stacking and/or van der Waals interactions between the *p*-methoxyphenyl ring and the second hydrophobic region of the MIF active site. It has been reported by us and other workers that the amino acids Pro-33, Tyr-36, Phe-49, Trp-108, and Phe-113 make up the second hydrophobic surface at the rim of the active site of MIF.^{10,14} These residues further contribute to the hydrophobic and hydrogen bond interactions between the pharmacophore and the active site of MIF.¹⁴ In further support of these interactions, we have previously shown that the L-amino acid Schiff base inhibitory effect was significantly improved by 5-fold upon changing the amino acid residue from L-phenylalanine (IC₅₀ = 50 μM) to L-tyrosine (IC₅₀ = 10 μM). This suggests that the increased potency was attributed to a hydrogen bond interaction within residues at the rim of MIF.¹⁰ Hydrazone **6** is 12 times more potent an inhibitor than L-tryptophan Schiff base **1**, so far the most potent inhibitor of MIF described in the literature.^{4,10–12,14,15}

Recently, we have discovered that monofluorination of **2** improved the inhibition of MIF activity.¹¹ As a consequence, we synthesized the 3-fluoro, 3-chloro, and 3-bromo-4-hydroxyphenyl derivatives of the more potent hydrazone, namely, the *p*-methoxyphenylhydrazone (**6**). The synthesis of the monohalogenated hydrazone derivatives involved treatment of a suspension of the 4-methoxyphenylhydrazine hydrochloride and the 3-halogenated 4-hydroxybenzaldehyde in methanol with aqueous sodium hydroxide (Table 1). A general increase in the inhibition of the MIF tautomerase activity was observed for the 3-halogenated 4-hydroxyphenylhydrazone derivatives **7–9** (Table 1). Among these hydrazones, **7** showed the most potent inhibition with an IC₅₀ of 130 nM while **8** and **9** gave values of 220 and 330 nM, respectively. The significant improvement in the inhibitory effect of these halogenated hydrazones **7–9** may be explained by the inductive effect that may lead to changes in the polarization of the hydroxyl moiety, thereby making it a stronger hydrogen bond donor/acceptor.

To determine if the activity would be affected by disubstitution on the nitrogen, **10** was synthesized (Figure 1). Methylation of phenylhydrazine (**5**) with methyl iodide and sodium amide afforded *N*-methyl-*N*-phenylhydrazine. Hydrazone formation under typical conditions gave **10** in 86% yield. To our disappointment **10** was a very weak inhibitor of MIF activity and displayed an IC₅₀ of 300 μM. This observation could be accounted for by increased steric hindrance or by the reduced number of hydrogen bonding interactions between the disubstituted hydrazone derivative **10** and the active site of MIF.

Table 2. Phenolic Hydrazone Carbamates and IC₅₀ for Inhibition of MIF Tautomerase Activity


Compounds ^a	Ar	R'	IC ₅₀ (μM) ^b
11		CH ₃	55
12		CH ₂ CH ₃	43
13		C(CH ₃) ₃	5.5
14			10
15			8

^a Compounds were characterized by ¹H NMR, ¹³C NMR, and MS (see Supporting Information). ^b Spectrophotometric analysis of MIF tautomerase activity on L-dopachrome methyl ester.

To further investigate the structure–activity relationship between the hydrazones and MIF, we looked at another class of compounds, namely, the phenolic hydrazone carbamates. The phenolic hydrazone carbamates **11–15** were chosen because they displayed key functionalities to **1** and **2** (Table 2). These functionalities are the hydroxyl group, phenyl ring, and the carboxylate moiety. It has been reported by us that a secondary hydrogen bond interaction exists between the carboxylate moiety of **2** and lysine-32A of the MIF active site.¹⁰ Significant improvement of approximately 10-fold in the inhibition of MIF tautomerase activity was observed on changing the methyl group of **11** (IC₅₀ = 55 μM) to the more lipophilic *tert*-butyl moiety **13** (IC₅₀ = 5.5 μM).¹¹ To determine if phenyl rings improve the inhibition of MIF activity, we replaced the *tert*-butyl carbamate with a benzyl carbamate **14** and *p*-methoxybenzyl carbamate **15**. This change from a bulky alkyl **13** to aromatic **14** and **15** (IC₅₀ = 10 and 8 μM, respectively) resulted in a slight decrease in the inhibitory effect.

Biological Activity. Intracellular MIF plays a critical role in mediating the cellular responses to pathways activated by LPS endotoxin,¹⁶ and MIF deficient cells are hyporesponsive to endotoxin.¹⁷ We and others have shown that MIF null macrophages can produce 50–60% less TNF compared to wild type.^{4,18} Previously, we have shown that anti-MIF antibody (10 μg/mL) inhibits 50% of TNF release from LPS-treated macrophages.⁴ We also have found that **2** dose-dependently inhibit LPS-induced TNF release from wild type and not MIF null macrophages, suggesting that the chemical inhibitor of MIF is specific. Accordingly, we reasoned that **7** binding to MIF would suppress LPS responses in macrophages. Compound **7** dose-dependently inhibited LPS-induced TNF release (Figure 2). Thus, **7** recapitulates the phenotype of the MIF deficient macrophages and is associated with decreased TNF production in response to LPS.

The importance of MIF as a molecular therapeutic target in sepsis has been confirmed by our recent observation that treatment with anti-MIF antibodies (4 mg/kg of monoclonal antibody against MIF (XIV.15.5) daily for 3 days) or **2** (40 mg/kg bidaily for 3 days) significantly improves survival in septic mice.⁴ We also recently discovered that serum MIF levels increased to 70% of maximum levels within 24 h after CLP and peaked at 36 h.⁴ This identified MIF as a late mediator in sepsis and indicated the therapeutic potential of inhibiting MIF

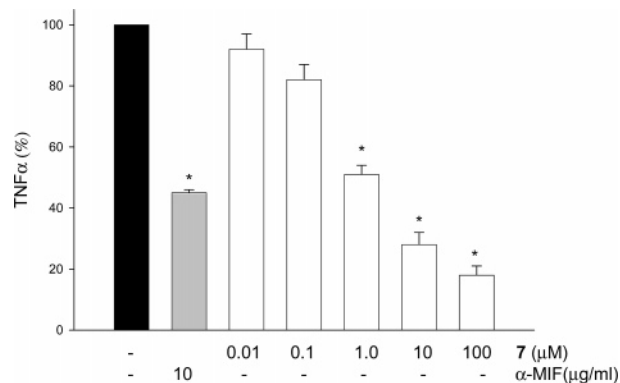


Figure 2. Compound **7** inhibits TNF secretion from LPS-treated macrophages. RAW 267.4 macrophages (10⁵) were treated with various concentrations of hydrazone **7** (0.01–100 μM) 30 min prior to LPS addition or with 10 μg/mL mouse monoclonal antibody against MIF (XIV.15.5, α-MIF). After 16 h of incubation, cell culture supernatants were collected for determination of TNFα concentration by ELISA. Data are presented as the mean ± SD (*n* = 3, (*) *p* < 0.01).

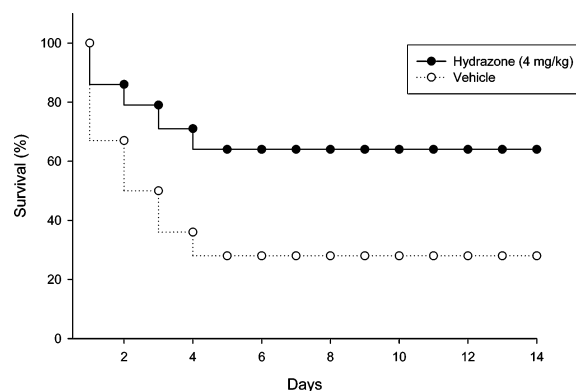


Figure 3. Compound **7** is protective after 24 h of late treatment in a CLP model. Mice were injected intraperitoneally with **7** (4 mg/kg) (*n* = 13, *p* < 0.01) or vehicle 24 h after CLP (*n* = 13). A single injection is composed of 100 μg of **7** (equivalent to 4 mg/kg) in 200 μL of 20% DMSO and 80% saline solution. Additional administrations of **7** (bidaily) were given on days 2 and 3.

in a clinically relevant time frame. Therefore, we reasoned that a delayed treatment with **7**, consistent with the kinetics of MIF release, could be successfully applied to improve survival in sepsis. We tested the ability of **7** to improve the survival rate in CLP-induced peritonitis, a widely used preclinical model of sepsis. Intraperitoneal treatment of **7** (4 mg/kg) initiated 24 h after CLP surgery and continued for 3 days resulted in a survival rate of 65% (*p* < 0.01) compared to 28% in the control (vehicle-treated) group (Figure 3). Thus, **7** treatment provided significant protection against sepsis lethality, comparable to the effect of anti-MIF antibody and **2**.⁴ Remarkably, a dose of **7**, 10-fold less than **2**, achieved similar protection. This finding indicates an association between the potency of **7** in inhibiting the MIF tautomerase active site and its beneficial effect of improving survival in experimental sepsis.

In summary, we have designed and synthesized phenolic hydrazones as nontoxic, potent MIF antagonists. Our structure–activity relationship study suggests that minor changes in functionalization of the hydrazones affect the binding of these compounds to the MIF active site. Notably, **7** exhibits the greatest activity of all the compounds tested and is so far the most potent inhibitor of MIF described in the literature.^{4,10–12,14,15} Compound **7** exhibited a potent anti-inflammatory activity *in vitro* as demonstrated by suppression of LPS-induced macrophage

age activation. Moreover, a relatively low concentration of this small-molecule MIF inhibitor improved survival in sepsis when treatment was initiated at 24 h after the onset of the disease.

Sepsis is a complex inflammatory disorder, and its clinical management is a challenging health issue. Therefore, our finding that **7** is effective at 24 h after the onset of the disorder could be of considerable clinical interest.

Experimental Section

General Procedure. All chemicals were obtained from commercial suppliers and used without further purification. Aluminum-backed silica gel 60 with 254 nm fluorescent indicator TLC plates were used. Developed TLC plates were visualized under a short-wave UV lamp, stained with an I₂-SiO₂ mixture. Flash column chromatography (FCC) was performed using flash silica gel (32–63 μm) and usually employed a stepwise solvent polarity gradient correlating with TLC mobility. Melting points were determined in a Gallenkamp melting point apparatus in open capillaries and are uncorrected. IR spectra were obtained on a Thermo Nicolet IR 100 FT-IR spectrometer. All ¹H spectra were recorded on a Joel spectrometer or a GE QE 300 spectrometer at 270 or 300 MHz. The ¹³C spectra were recorded on a GE QE 300 spectrometer at 75 MHz. Chemical shifts are relative to the deuterated solvent peak and are in ppm. The coupling constants (*J*) are measured in Hz. The ¹H signals are described as s (singlet), d (doublet), t (triplet), q (quartet), m (multipet), or br s (broad singlet). Low- and high-resolution mass spectrometry was carried out at the Mass Spectrometry Facility at the University of Illinois at Urbana-Champaign.

General Procedure for Compounds 3–5 and 10–15. 4-Hydroxybenzaldehyde (122 mg, 1 mmol) or 3-fluoro-4-hydroxybenzaldehyde (140 mg, 1 mmol) and the hydrazide (2 mmol) were dissolved in ethanol (10 mL). To this was added acetic acid (1 mmol), and the mixture was stirred overnight at room temperature. Removal of the ethanol in vacuo afforded an oily residue. The residue was taken up with ethyl acetate and washed with water. The organic layer was separated and dried with anhydrous Na₂SO₄. Concentration in vacuo afforded a residue that was subsequently purified by FCC using hexane and ethyl acetate as eluent (4:1) to give **3–5** and **10–15**.

General Procedure for Compounds 6–9. 4-Hydroxybenzaldehyde (122 mg, 1 mmol) or 3-fluoro-4-hydroxybenzaldehyde (140 mg, 1 mmol) or 3-chloro-4-hydroxybenzaldehyde (156 mg, 1 mmol) or 3-bromo-4-hydroxybenzaldehyde (201 mg, 1 mmol) and 4-methoxyphenylhydrazine hydrochloride (350 mg, 2 mmol) were suspended in methanol (10 mL). To this suspension was added a 2 M aqueous solution of sodium hydroxide (60 mg, 1.5 mmol), and the mixture was stirred overnight at room temperature. Upon completion of the reaction, the solution was then acidified to pH 4 by the addition of 1 M HCl. Removal of the methanol in vacuo afforded an oily residue. The residue was taken up with ethyl acetate and washed with water. The organic layer was separated and dried with anhydrous Na₂SO₄. Concentration in vacuo afforded a residue that was subsequently purified by FCC using hexane and ethyl acetate as eluent (4:1) to give **6–9**.

Spectrophotometric Assay for Enzymatic Activity. A fresh stock solution of L-dopachrome methyl ester (2.4 nM) was generated by oxidation of L-3,4-dihydroxyphenylalanine methyl ester with sodium periodate, producing an orange solution. Activity was determined at room temperature by adding dopachrome methyl ester (0.3 mL) to a cuvette containing 1 μL of MIF solution (850 ng/mL) in 50 mM potassium phosphate buffer, pH 6, and measuring the decrease in absorbance from 2 to 20 s at 475 nm spectrophotometrically. Inhibitors **3–15** were dissolved in DMSO at various concentrations (0.1–100 μM), and 1 μL was added to the cuvette with the MIF prior to the addition of the dopachrome.

Cellular Assay. Compound **7** inhibits TNF secretion from LPS-treated macrophages. RAW 267.4 macrophages (10⁵) were treated with various concentrations of hydrazone **7** (0.01–100 μM) 30 min prior to LPS addition. After 16 h of incubation, cell culture

supernatants were collected for determination of TNF concentration by ELISA. Data are presented as the mean ± SD (*n* = 3, *p* < 0.01).

Animal Studies. All animal experiments were approved by the Institutional Animal Care and Use Committee of the North Shore-Long Island Jewish Research Institute. Male Balb/C mice, ~8 weeks old, were subjected to cecal ligation and puncture. Details of the CLP procedure has been carried out as follows: In anesthetized male BALB/c mice (100 mg/kg ketamine and 8 mg/kg xylazine administered intramuscularly) the cecum was ligated and given a single puncture. Abdominal access was gained via a midline incision. The cecum was isolated and ligated with a 6-0 silk suture below the ileocecal valve, and the cecum was punctured once with a 22G needle. Stool (approximately 1 mm) extruded from the hole, and the cecum was placed back into the abdominal cavity. The abdomen was closed with two layers of 6-0 Ethilon sutures. Antibiotics were administered immediately after CLP (0.5 mg/kg Premaxin, subcutaneously, in a total volume of 0.5 mL/mouse), and a single dose of resuscitative fluid (normal saline solution) was administered subcutaneously (20 mL/kg body weight) immediately after CLP surgery.¹⁹ Mice were injected intraperitoneally with 4 mg/kg (*n* = 13, *p* < 0.01) or vehicle 24 h after CLP (*n* = 13). Two additional injections were given on days 2 and 3. Vehicle (aqueous 20% DMSO) or **7** (4 mg/kg, intraperitoneally) treatment was started 24 h after the induction of sepsis and repeated twice daily on days 2 and 3. Animal survival was monitored for 14 days.

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Supporting Information Available: Physical and spectral data for **3–15**. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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